



## Comparison of Chemical Composition and Antioxidant Potential of Hydrodistilled Oil and Supercritical Fluid CO<sub>2</sub> Extract of *Valeriana wallichii* DC

D. Sugumar Pandian, N.S. Nagarajan\*

Department of Chemistry, Gandhigram Rural Institute, Deemed University, Gandhigram – 624 302, TN, India.

### ARTICLE DETAILS

#### Article history:

Received 01 October 2015

Accepted 12 October 2015

Available online 23 October 2015

#### Keywords:

*Valeriana wallichii* DC

Hydrodistillation

Supercritical Fluid CO<sub>2</sub> Extraction

Antioxidant Activities

GC-MS

### ABSTRACT

In the present study, chemical constituents of the essential oil and supercritical fluid extracts of *Valeriana wallichii* DC obtained by hydrodistillation and supercritical fluid CO<sub>2</sub> extraction have been studied by GC-MS technique. Seventy six compounds were obtained from hydro distillation, representing 98.6% of the total oil. The major component of hydro distillation was isovaleric acid (37.69%), methylvaleric acid (17.24%) and seychellene (6.49%) along with many other components in minor amounts. Ninety eight and seventy six components were identified by GC-MS in the supercritical CO<sub>2</sub> fluid extracts extracted at 100 and 200 bar respectively, where the major components were cis-adamantane-2-carboxylic acid, 4-hydroxy (36.88%, 40.26%), isovaleric acid (5.91%, 6.75%), β-bisabolol (6.48%, 5.71%), bornyl isovalerate (5.08%, 5.54%), of the *Nardostachys jatamansi* extract at 200 and 100 bar respectively. Other major components present in the supercritical fluid extracts were β-methasone valerate, valeranone, benzyl isovalerate, nerolidol, methylvaleric acid and geranyl isovalerate which were usually absent or trace in the hydrodistilled essential oil. The antioxidant activity of essential oil and supercritical fluid extracts was evaluated in DPPH radical, superoxide radical, hydroxyl scavenging assays. Reducing power of those oil and extracts were also studied. Essential oil and Supercritical fluid extracts gave comparable DPPH, superoxide radical scavenging and reducing power activity and there is no much deviation among various activities was observed. However, when we consider overall antioxidant assays, 200 bar extract of valerian is the best among essential oil and extracts.

### 1. Introduction

Indian valerian roots of *Valeriana wallichii* de Candolle, family *Valerianaceae*, which grows wild in the temperate Himalayas on the borders of Kashmir at altitude of 1500-3000 m and Afghanistan [1]. It is commonly known as 'tagar' in India and also called as Valerian, Indian valerian, Valerian jatamansi [2]. It is an ingredient of herbal medicines in Indian systems of medicine [3]. It is a small perennial herb of 14-45 cm height, with root stock, thick branching stem, sharply pointed leaves, white or pink flowers in clusters and hairy fruit [4]. The rhizome is solid also tough internally, it is greenish-brown in colour along with the odour is effectively valerianaceous [5].

Roots of *V. wallichii* are used as an aphrodisiac, insecticide and in mental disorders [6]. In India, Valeriana has long been used in Ayurveda and Unani systems of medicine, which describes its uses in skin diseases, Insanity, epilepsy and snake bite and considered to have remarkable sedative effects in nervous unrest, stress and neuralgia [7]. Valerian is the top ten selling retail herb for herbal supplements in North America and Australia [8]. It has also been prescribed as the perfect herbal tranquilizer, and was used for this purpose in the First World War to treat soldiers suffering from shell shock [9]. Antiinflammatory [4], antispasmodic [10], antioxidant [11], larvicidal [12], antianxiety [13], anti HIV [14], anti-diarrhoeal, and bronchodilatory activities [15] of Valeriana have been scientifically reported. The plant is also used as cytotoxic [16]. Its essential oil exhibited antimicrobial activity against pathogenic bacteria and also exhibited potent antifungal activity against different human and plant fungal pathogens [17].

Literature survey revealed the presence of flavone glycosides [18-19] iridoids and lignans [20-22] in *V. wallichii*. The essential oil from root contain calarene, α-santalene, α-curumene, xanthorrhizol, valeranone, α, β and γ-patchoulene, α-fenchene, patchouli alcohol, maaliol, β-sitosterol,

maali-oxide, valerenic acid, isovaleric and β-methylvaleric acid, formic, propionic, butyric, palmitic acid and stearic acids, and isovaleryl ester of D-α-hydroxyisovaleric acid. Patchouli alcohol (40%) as the major constituents followed by the presence of δ-guaiene (10%), seychellene (8%), acetoxyl patchouli alcohol (5%). The presence of valepotriates like valtrate, isovalerohydroxy-dihydrovaltrate, 1α-acevaltrate, and didrovaltrate [23]. The root rhizome parts are highly aromatic and contain valepotriates and essential oils. The essential oil showed maaliol (36.8%) as the major constituent followed by the presence of β-gurjunene (21.3%), acoradiene (9.9%), guaiol (8.6%) and α-santalene (5.5%) [24].

There are few studies published about the chemical composition and pharmacological aspects of *Valeriana wallichii* and almost all of these published essential oil composition were different from each other. Therefore, the aim of study is to determine the chemical composition of the essential oil and compare with chemical composition of supercritical fluid extracts of *Valeriana wallichii*. However, to the best of our knowledge, this is the first report on the gas chromatography-mass spectrometry (GC-MS) to study the chemical composition of supercritical fluid extracts of *Valeriana wallichii*. Hence, in the present investigation was made to study the antioxidant potential of *Valeriana wallichii* essential oil and supercritical fluid extracts and chemical constituents of the same were analysed by GC-MS.

### 2. Experimental

#### 2.1 Plant Material

The dried rhizomes of *Valeriana wallichii* DC were purchased from the commercial market and the plant was authenticated by Dr. R. Kumuthakalavalli (Department of Biology, Gandhigram Rural Institute, Deemed University, Gandhigram, Tamil Nadu, India) and a voucher specimen is kept in the department of chemistry of the University for future reference.

\*Corresponding Author

Email Address: nsnrajan@yahoo.co.in (N.S. Nagarajan)

## 2.2 Chemicals

1-Diphenyl-2-picrylhydrazyl (DPPH) radical, ascorbic acid, potassium ferricyanide, trichloroacetic acid (TCA), hydrogen peroxide and phosphate buffer (pH 7.4), methanol, distilled Water, CO<sub>2</sub>,  $\alpha$ -tocopherol, tris-HCl buffer, nitroblue tetrazolium chloride (NBT), butylated hydroxytoluene (BHT), ethylene diamine tetraacetic acid (EDTA), thiobarbituric acid (TBA), nicotinamide adenine dinucleotide (NADH), deoxyribose, ferric chloride. All chemicals used including solvents were of analytical grade.

## 2.3 Isolation of Oil and Extract

### 2.3.1 Hydrodistillation Method

The classical method of hydrodistillation using the Clevenger type apparatus for 4 h was used for the isolation of the essential oil from *Valeriana wallichii*. The essential oil was collected and stored at 4 °C until their analysis by gas chromatography/mass spectrometry (GC-MS) and other studies. The essential oil yield was 4 %.

### 2.3.2 Supercritical Fluid CO<sub>2</sub> Extraction Method

Supercritical CO<sub>2</sub> extraction was performed on a supercritical fluid extractor (ATI technology, Bangalore, India) with the extractor volume 5 kg. The flow rate of CO<sub>2</sub>, the extraction temperature and pressure were adjusted by the control panel, and the extraction time was measured by the stopwatch. The CO<sub>2</sub> gas supplied from storage tank to super cooler, where gas CO<sub>2</sub> converted into liquid CO<sub>2</sub> and it went to the extractor. Before liquid CO<sub>2</sub> passed into the extraction vessel, filled with the samples, by the means of a pump, it was pressurized to the desired pressure and heated to the specified temperature in order to reach the supercritical state. In this study, extractions were performed at temperatures 55 °C, two different pressure levels (100 and 200 bar) and dynamic extraction time 1.5 hours each. The supercritical CO<sub>2</sub> flow rate was maintained at 2.5 mL/min and the duration of static extraction time was fixed to 55 °C min. The introduction of some rigid materials such as glass beads with the ground sample, contributed to maintaining a proper flow rate of CO<sub>2</sub> in the extractor vessel as well as in maintaining the desired permissibility of the particle during extraction process [25-26]. The powdered plant material (2 kg) placed into the extractor vessel. The extractions were performed, during the dynamic extraction time, CO<sub>2</sub> carrying the crude extract flowed out of the extraction vessel unit and into a collection vessel. The extracts for each parameter were stored in amber coloured tubes in a refrigerator (4 °C) until further analysis and the yields were 0.41 kg at 100 bar and 0.45 kg at 200 bar. The identification of extracted constituents was carried out by GC-MS.

### 2.4 GC-MS Analysis

The oil and CO<sub>2</sub> extract was analyzed by GC-MS using an Agilent GC 6890N model gas chromatograph-5973N model mass spectrometer equipped with a 7683B series auto-sampler injector (Agilent, USA). The GC was equipped with a HP-5MS capillary column (30 m x 0.25 mm x 0.25  $\mu$ m film thickness). The GC oven temperature was kept at 40 °C for 3 min and programmed to 100 °C at a rate of 25 °C for 5 min, 180 °C at a rate of 25 °C for 5 min, 220 °C at a rate of 25 °C for 5 min and 280 °C at a rate of 25 °C for 12 min. Both oil and extract was analyzed by dissolving in CHCl<sub>3</sub>. Injection volume was 1.0  $\mu$ L, inlet pressure was 7.06 psi and the injector temperature was 250 °C. Helium was used as carrier gas and linear velocity (*u*) was 36 cm/sec. Injection mode was split (30:1) and MS interface temperature was 250 °C. Mass spectra were recorded in the scan mode at energy was 70 eV and MS spectra were scanned from 50 to 550 m/z at 2.2 scan s<sup>-1</sup>. Compound identification was based on the comparison of estimation retention indices using a MS library. The NIST 05a and Wiley7a spectrometer data bank was used to determine the percentage composition of the compounds [27].

### 2.5 DPPH Radical Scavenging Activity

Various concentration of hydrodistilled and supercritical fluid CO<sub>2</sub> extracted samples of *Valeriana wallichii* roots individually (0.3 mL of hydro distilled oil, CO<sub>2</sub> extract at 100 bar and 200 bar) mixed with 2.7 mL of methanol solution containing DPPH radicals (6 $\times$ 10<sup>-5</sup> mol/L). The mixture was shaken vigorously and allowed to stand for 60 minutes in the dark. The reduction of the DPPH radical was determined by reading the absorbance at 517 nm. Ascorbic acid and  $\alpha$ -tocopherol were used as standards [28]. The radical scavenging activity (RSA) was calculated as a percentage of DPPH discoloration, using the formula;

$$\% \text{ RSA} = \frac{(\text{Abs}_{\text{DPPH}} - \text{Abs}_s)}{(\text{Abs}_{\text{DPPH}})} \times 100$$

Where Abs<sub>s</sub> is the absorbance of the extract sample and Abs<sub>DPPH</sub> is the absorbance of the DPPH solution.

## 2.6 Superoxide Radical Scavenging Activity

The superoxide radical scavenging activity of the various concentrations of hydrodistilled and supercritical fluid CO<sub>2</sub> extracted samples of *Valeriana wallichii* was studied using the method [29]. Superoxide radicals are generated by 1 mL of Tris-HCl buffer (16 mM, pH=8), 1 mL of NBT (50  $\mu$ M), 1 mL NADH (78  $\mu$ M). The reaction mixture was incubated at 25 °C for 5 min and the absorbance was measured at 560 nm (Jenway 6100, Dunmow, Essex, UK). A control tube containing Tris-HCl buffer was also processed in the same way without test sample. Different concentration of ascorbic acid and BHT were used as standards. The radical scavenging activity (RSA) was calculated as a percentage of superoxide discoloration, using the equation: % RSA = [(A<sub>control</sub> - A<sub>sample</sub>) / A<sub>control</sub>] x 100.

## 2.7 Hydroxyl Radical Scavenging Activity

The hydroxyl radical scavenging activity of the various concentrations of hydro distilled and supercritical fluid CO<sub>2</sub> extracted samples of *Valeriana wallichii* was estimated by following the method [30]. The hydroxyl radical was generated by a fenton-type reaction. The reaction mixture contained 0.2 mL of sample in varied concentrations to which, 0.1 mL EDTA (1 mM) -FeCl<sub>3</sub> (10 mM) mixture, 0.1 mL H<sub>2</sub>O<sub>2</sub> (10 mM), 0.36 mL deoxyribose (10 mM), 0.33 mL phosphate buffer (50 mM, pH 7.4) and 0.1 mL of ascorbic acid (1 mM) was added in sequence. The mixture was incubated at 37 °C for 1 h. To this mixture was added 1.0 mL each of TCA (10%) and TBA (0.67%) and kept in boiling water bath for 20 minutes. The colour developed was read at 532 nm. The control tube contains phosphate buffer, instead of sample.

## 2.8 Reducing Power

The reducing power of *Valeriana wallichii* was determined [31]. Various concentration of hydrodistilled and supercritical fluid CO<sub>2</sub> extracted samples of *Nardostachys jatamansi* DC of (1 mL), phosphate buffer (1 mL, 0.2M, pH=6.6) and potassium ferricyanide (1 mL, 10 mg/mL) were mixed together and incubated at 50 °C for 20 min. TCA (1 mL, 100 mg/mL) was added to mixture and centrifuged at 8,000 rpm for 5 min. The supernatant (1 mL) was mixed with distilled water (1 mL) and ferric chloride (0.1 mL, 1 mg/mL) and then the absorbance was measured at 700 nm.

## 2.9 Statistical Analysis

All the results are expressed as mean values standard deviation (SD), n = 3. Data were analyzed using one way analysis of variance (ANOVA) followed by Tukey's multiple comparison *post hoc* test using SPSS software 16.0 versions. Values of *p* < 0.05 were considered statistically significant [32].

## 3. Results and Discussion

### 3.1 Chemical Composition of the Extract and Essential Oil

The *Valeriana wallichii* DC extract were extracted in supercritical fluid CO<sub>2</sub> extraction method. Ninety eight (100 bar) and seventy six (200 bar) compounds were identified, representing, 98.6% and 98.76% respectively of the total extract as shown in Table 1 by GC-MS analysis. Their main compounds were cis-adamantane-2-carboxylic acid, 4-hydroxy (36.88%, 40.26%), isovaleric acid (5.91%, 6.75%),  $\beta$ -bisabolol (6.48%, 5.71%), bornyl isovalerate (5.08%, 5.54%), isoquinolin-6,7-diol-1-carboxylic acid, N-acetyl-1-methyl- (1.74%, 3.87%), patchouli alcohol (2.19%, 2.22%),  $\beta$ -methasone valerate (1.33%, 2.34%), 1H-indole, 1-acetyl-2,3-dihydro-6-nitro (1.50%, 1.49%), valtrate (0.92%, 1.80%), benzyl isovalerate (0.96%, 1.45%), nerolidol (1.64%, 0.83%), valeranone (1.64%, 0.63%), methylvaleric acid (1.15%, 1.19%), geranyl isovalerate (0.91%, 0.80%) at 100 and 200 bar respectively. Some of the compound only found in CO<sub>2</sub> extract at 100 bar which were propyl valerate, p-cresol, citronellol, phenyl ethyl alcohol and bornyl butyrate. At the same time some of the compound only found in CO<sub>2</sub> extract at 200 bar which were 2,6-Diamino-4-cyclohexyl-4H-thiopyran-3,5-dicarbonitrile and cholestan-3-one, cyclic 1,2-ethanediyol aetal. The phenolic compound of such as 4-terpineol,  $\alpha$ -terpineol, carvacrol, p-cresol, patchouli alcohol and  $\alpha$ -tocopherol were present in both extracts. The fatty compounds present in 100 and 200 bar extract were palmitic acid, eicosane, tricosane, tetracosane and methyl linoleate etc.

The essential oil of *Valeriana wallichii* was extracted for hydrodistillation method. The results obtained by GC-MS analysis of the oil are presented in Table-1. Seventy six compound of the total oil were identified, representing 98.6%. The major compounds were isovaleric acid (37.69%), methylvaleric acid (17.24%) and seychellene (6.49%). Other compounds

were present considerable amount such as calarene (3.24%),  $\gamma$ -gurjunene (3.40%), patchouli alcohol (3%), aromadendrene (1.62%),  $\alpha$ -patchoulene (2.03%),  $\beta$ -patchoulane (1.50%).

**Table 1** Chemical constituents and retention indices of the *V. wallichii* oil, SFE extract 100 and 200 bar

Componentes	CO <sub>2</sub>	CO <sub>2</sub>	H.oil	RI					
	Extract	Extract							
	100 bar	200 bar							
l-Propyl 6,9,12-hexadecatrienoate	0.21	--	--	203					
Methyl isovalerate	--	0.01	0.13	721					
Isovaleric acid	5.91	6.75	37.69	811					
Furfural	--	--	0.21	831					
2-Butenoic acid, 3-methyl-	--	--	0.15	860					
Ethyl valerate	--	--	0.03	884					
Methylvaleric acid	1.15	1.19	17.24	910					
(-)- $\beta$ -Pinene	0.10	0.19	--	943					
Camphene	0.64	0.77	0.12	943					
$\alpha$ -Pinene	0.13	0.24	--	948					
Propyl valerate	0.03	--	--	984					
cis-2,6-Dimethyl-2,6-octadiene	0.56	0.21	--	985					
$\gamma$ -Terpinene	0.01	0.01	--	998					
Limonene	0.06	0.08	0.06	1018					
Cycloheptane, 1,3,5-tris(methylene)-	0.10	--	--	1039					
O-Cymene	0.02	0.03	0.06	1042					
Eucalyptol	--	--	0.01	1059					
Linalool	0.06	0.02	0.27	1082					
Phenylethyl Alcohol	0.01	--	--	1136					
4-Terpineol	0.11	0.13	--	1137					
Borneol	0.34	0.17	0.04	1138					
$\alpha$ -Terpineol	0.05	0.03	0.04	1143					
cis-Linalool oxide	--	--	0.05	1164					
1,2-Diisopropylbenzene	--	--	0.38	1176					
Citronellol	0.03	--	--	1179					
Amyl valerate	--	--	0.01	1183					
P-Creosol	0.04	--	--	1203					
Cyclolongifolene oxide, dehydro-	--	--	0.19	1208					
$\alpha$ -Santalene	0.51	0.20	0.69	1211					
Hexyl isovalerate	0.06	--	0.05	1218					
$\alpha$ -Copaene	0.04	--	0.03	1221					
Geraniol	0.08	0.08	--	1228					
Thymol methyl ether	0.15	0.14	0.08	1231					
Furfuryl 3-methylbutanoate	--	--	0.12	1243					
Carvacrol	0.66	0.02	--	1262					
1,2,6-Hexanetriol	--	--	0.02	1265					
Seychellene	0.73	0.35	6.49	1275					
Bornyl acetate	0.92	0.35	0.10	1277					
Isobornyl acetate	0.20	0.24	--	1277					
2 $\alpha$ , 4 $\alpha$ $\beta$ , 8 $\alpha$ $\beta$ -Decahydro-2-naphthalenol	--	0.12	--	1289					
Ledene oxide-(ii)	--	--	0.15	1293					
Neoclovene oxide	--	--	--	1305					
Megastigma-4,6(e),8(z)-triene	--	--	0.64	1317					
Geranyl acetate	0.47	0.18	--	1352					
Ethanone, 1-(2,4,5-trimethylphenyl)-	--	--	0.23	1369					
$\alpha$ -Copaen-11-ol	--	--	0.30	1377					
Ledane	--	--	0.17	1380					
7-Tetracyclo[6.2.1.0(3.8)0(3.9)]undecanol, 4,4,11,11-tetramethyl	--	--	2.55	1385					
3-Iodomethyl-3,6,6-trimethyl-cyclohexene	--	0.12	--	1385					
Alloaromadendrene	--	--	0.64	1386					
Aromadendrene	0.12	--	1.62	1386					
Benzene, 2-(1,1-dimethylethyl)-1,4-dimethoxy-	0.15	--	--	1386					
Patchoulane	--	--	0.21	1393					
$\beta$ -Patchoulane	--	--	1.50	1393					
Benzyl isovalerate	0.96	1.45	--	1394					
Aromadendrene, dehydro-	--	--	0.92	1396					
Isolongifolene, 9,10-dehydro-	--	--	0.45	1398					
Neoisolongifolene, 8,9-dehydro-	--	--	0.50	1398					
$\alpha$ -Patchoulene	0.21	0.10	2.03	1403					
Calarene	--	--	3.24	1403					
(-)- $\alpha$ -Panasinsen	0.19	--	0.71	1416					
Widdrene	--	--	0.08	1416					
Patchouli alcohol	2.19	2.22	3.00	1420					
$\alpha$ -Bergamotene	0.36	0.15	0.27	1430					
Cyperene	0.17	--	--	1432					
1,1,4a-Trimethyl-5,6-dimethylenedecahydronaphthalene	--	--		0.10	1443				
$\alpha$ -Farnesene	0.11	0.15	--	1458					
$\gamma$ -Gurjunene	--	--		3.40	1461				
Aromadendrene oxide-(2)	--	--		0.97	1462				
$\gamma$ -Elemene	0.10	--		0.08	1465				
$\beta$ -Selinene	--	--		1.00	1469				
Eudesma-4(14),11-diene	--	--		0.12	1469				
Eremophilene	--	--		0.10	1474				
Valencene	--	--		0.17	1474				
$\alpha$ -Selinene	--	--		0.21	1474				
Bornyl butyrate	0.61	--		0.16	1476				
cis-Adamantane-2-carboxylic acid, 4-hydroxy-	36.88	40.26	--	1486					
$\alpha$ -Guaiene	0.41	0.15		0.13	1490				
$\delta$ -Guaiene	1.39	0.45		1.02	1490				
Phenethyl isovalerate	0.80	0.46	--	1493					
Caryophyllene	0.07			0.05	1494				
Bicyclo[7.2.0]undec-4-ene, 4,11,11-trimethyl-8-methylene	1.42	0.42	--	1494					
Germacrene A	0.78	0.45	0.29	1505					
Caryophyllene oxide	0.18	0.18	0.27	1507					
Eudesma-3,7(11)-diene	--	0.09	--	1507					
Bornyl Isovalerate	5.08	5.54	--	1512					
cis- $\alpha$ -Bisabolene	0.55	0.19	--	1518					
B-Guaiene	0.92	--	--	1523					
$\alpha$ -Curcumene	0.98	0.35	--	1524					
Caryophyllene-(i1)	--	--		1.03	1528				
Epiglobulol	0.17	0.10		0.49	1530				
Spathulenol	-	--		0.70	1536				
1-Isopropenyl-3,3-dimethyl-5-(3-methyl-1-oxo-2-butenyl)cyclopentane	0.38	--	--	1541					
Limonen-6-ol, pivalate	0.40	0.37	--	1560					
Nerolidol	1.64	0.83	--	1564					
$\beta$ -Humulene	--	--		0.09	1574				
Bornyl valerate	--	--		0.75	1576				
Geranyl isovalerate	0.91	0.80		1586					
2-Butenal, 2-methyl-4-(2,6,6-trimethyl-1-cyclohexen-1-yl)-	--	--		0.56	1586				
Isolongifolen-5-one	0.19	--	--	1587					
Ethanone, 1-[2-(5-hydroxy-1,1-dimethylhexyl)-3-methyl-2-cyclopropen-1-yl]-	0.73	0.57	--	1591					
Valeranone	1.64	0.63	--	1615					
$\beta$ -Bisabolol	6.48	5.71	--	1619					
1(2h)-Naphthalenone, octahydro-4,8a-dimethyl-6-(1-methylethenyl)-, (4 $\alpha$ ,4 $\alpha$ $\beta$ ,6 $\alpha$ ,8 $\alpha$ $\beta$ )-	0.38	0.48	0.43	1634					
Cedren-13-ol, 8-	--	--		0.17	1646				
Baldrial	0.69	0.90	--	1689					
6,7-Dimethoxy-2-tetralone	0.64	0.63	--	1716					
Murolan-3,9(11)-diene-10-peroxy	--	--		0.21	1729				
Khusilic acid	--	--		0.20	1747				
Phenol, 2-methyl-5-(1,2,2-trimethylcyclopentyl)-, (s)-	0.31	0.27	--	1776					
1H-Indole, 1-acetyl-2,3-dihydro-6-nitro-	1.50	1.49	--	1791					
Ethyl 9,9-Diformylnona-2,4,6,8-tetraenoate	0.19	0.71	--	1867					
Methyl palmitate	--	--		0.11	1878				
Bicyclo[4.4.0]dec-2-ene-4-ol, 2-methyl-9-(prop-1-en-3-ol-2-yl)-	0.06	0.52	0.98	1904					
6-(1-Hydroxymethylvinyl)-4,8a-dimethyl-3,5,6,7,8,8a-hexahydro-1h-naphthalen-2-one	--	--		0.77	1916				
Cyclohexadecane	0.12	--	--	1918					
Bicyclo[4.4.0]dec-5-ene, 1,5-dimethyl-3-hydroxy-8-(1-methylene-2-hydroxyethyl-1)-	--	--		0.30	1933				
Estra-1,3,5(10)-trien-17 $\beta$ -ol	0.59	0.43	--	1949					
Coumarin-6-ol, 3,4-dihydro-5,7,8-trimethyl-	0.95	0.97	--	1952					
2,6-Dimethyl-8-(tetrahydropyran-2-yloxy)-octa-2,6-dien-1-ol	0.46	0.85	--	1953					
Palmitic acid	--	0.40	--	1968					
Methyl (10E)-10-heptadecen-8-ynoate	0.06	--	--	2003					
Eicosane	1	0.14	--	2009					
Deoxysericealactone	0.19	0.09	--	2025					
9-Methyl-Z,Z-10,12-hexadecadien-1-ol acetate	--	0.07	--	2029					
Methyl linoleate	0.13	--	--	2093					
Methyl 9,12,15-octadecatrienoate	0.21	--	--	2101					
Methyl 10,13-octadecadiynoate	0.04	--	--	2112					
Valeric acid, 4-pentadecyl ester	--	--		0.02	2112				
Androstan-17-one, 3-ethyl-3-hydroxy-, (5 $\alpha$ )-	0.12	--	--	2251					
Tricosane	0.23	--	--	2307					

9-Tricosene, (z)-	0.35	--	--	2315
14-Oxononadec-10-enoic acid, methyl ester	0.80	--	--	2321
2-(5-Hydroxypent-2-ynyl)-3-oxocyclopentylthioacetic acid, s-t-butyl ester	0.58	0.66	--	2350
Tetracosane	0.16	--	--	2407
Isoquinolin-6,7-diol-1-carboxylic acid, N-Acetyl-1-methyl-	1.74	3.87	--	2447
2h-Pyran, 2-(7-heptadecyloxy)tetrahydro-	0.19	0.06	--	2453
1,3,5-Cycloheptatriene, 2,5-bis(Tetrahydropranyloxymethyl)-7,7-dimethyl-	0.84	0.92	--	2549
2,6-Diamino-4-cyclohexyl-4H-thiopyran-3,5-dicarbonitrile	--	1.69	--	2582
Chiapin b	--	--	0.13	2612
Cholestan-3-ol, 2-methylene-, (3 $\beta$ ,5 $\alpha$ )-	0.37	0.99	0.12	2652
Valtrate	0.92	1.80	--	2686
Glyceryl linolenate	0.21	0.69	--	2705
$\gamma$ -Sitosterol	0.12	0.37	--	2731
Cholestan-3-one, cyclic 1,2-ethanediy aetal, (5 $\beta$ )-	--	0.56	--	2770
Nonacosane	0.38	--	--	2904
Squalene	0.28	0.32	--	2914
Clocortolone Pivalate	0.50	1.14	--	3124
$\alpha$ -Tocopherol	0.05	0.27	--	3149
$\beta$ - Methasone valerate	1.33	2.34	--	3307
1-Heptatriacotanol	0.32	0.27	--	3942
3',8,8'-Trimethoxy-3-piperidyl-2,2'-binaphthalene-1,1',4,4'-tetrone	1.04	1.61	--	4187

It was reported hydrodistilled oil contains valerianian,  $\alpha$ -pinene, camphene and terpineol [33-34]. Other constituents present in roots were  $\alpha$ -valene,  $\beta$ -bisabolene,  $\beta$ -elemene,  $\beta$ -phellandrene,  $\beta$ -pinene,  $\beta$ -valene, borneol, bornyl acetate, bornylformate, camphene, limonene, myrcene, caryophyllene. A study revealed that the chemical compositions of the oil show two chemotypes within *V. wallichii*. The type 1<sup>st</sup> was characterized by presence of maaliol (64.3%), viridiflorol (7.2%) and sesquiterpene hydrocarbons (19.2%). The type 2<sup>nd</sup> contained patchouli alcohol (40.2%), viridiflorol (5.2%), acetoxy-patchouli alcohol (4.5%) and sesquiterpene hydrocarbons (34.5%) [35-36]. Two new flavone glycosides, acacetin 7-O- $\beta$ -sophoroside and acacetin 7-O-(6-O- $\alpha$ -l-rhamnopyranosyl)- $\beta$ -sophoroside were isolated from the rhizomes and roots of *V. wallichii* [37]. Rhizomes and roots contain cyclopentapyrans, acacetin-7-O-rutinosides, linarin-iso-valerinate, 4-methoxy-8-pentyl-1-naphthoic acid [38], lignin prinsepiol-4-omicron- $\beta$ -d-glucoside, coniferin, hexacosanic acid, limonene, choline, chatinine, valerianine, actinidine, tannins and resins 4-methoxy, 8-pentyl-1-naphthoic acid, methyleicosanoate, cubenol, caryophyllene oxide, cadinol and aristolene are other constituents isolated from this plant [39-40].

The aroma compounds were present tiny amount in CO<sub>2</sub> extract as well as oil such as furfural, ethyl valerate,  $\alpha$  and  $\beta$  pinene, limonene, cymenecitronellol, geraniol, phenyl ethyl alcohol, linalool, bornyl acetate, linalool oxide, hexyl isovalerate, amyl valerate etc.

To our knowledge, this is the first report on the composition of supercritical fluid CO<sub>2</sub> extract of *Valeriana wallichii* DC. The new compound identified from the extract of *Valeriana wallichii* DC were chiapin b and clocortolone pivalate in very small amount.

### 3.2 DPPH Radical Scavenging Activity

Percent DPPH radical scavenging capability of hydrodistilled *Valeriana wallichii* essential oil along with supercritical fluid CO<sub>2</sub> against the concentration was illustrated in Fig. 1. The scavenging effect of the essential oil (40.25% - 60.54%), supercritical fluid extract at 100 bar (30.27 - 52.57%) and supercritical fluid extract at 200 bar (39.57% - 63.05%) on DPPH radical linearly increased with increasing concentration from 0.2 to 1.0 mg. All the essential oil and supercritical fluid extract showed good to excellent percent scavenging activity in comparison with BHA (47.38% - 89.39%) and Ascorbic acid (56.96% - 90.67%). At the higher concentration (1 mg) *V. wallichii* supercritical fluid extracts are able to give equally good radical scavenging activity in comparison with positive control ascorbic acid. At lower concentration (0.2 mg) *Valeriana wallichii* essential oil gave better DPPH radical activity than supercritical fluid extracts. Based on IC<sub>50</sub> value, the percent radical scavenging activity can be ranked in the following order, Hydrodistilled *V. wallichii* essential oil (0.51 mg/mL) > SFE of *V. wallichii* 200 bar (0.53 mg/mL) > SFE of *V. wallichii* 100 bar (0.92 mg/mL). Sudhanshu et al. (2012) reported methanolic extract of *V. wallichii* showed appreciable DPPH scavenging activity which is comparable to standard ascorbic acid. Our results are well

supported with the above literature. It has been reported that presence of phenolic compounds plays an important role in antioxidant activity of natural products [41]. From GC-MS studies (Table 1), we can see supercritical fluid extract at 100 bar contains 0.04% p-cresol, 0.66% of carvacrol and 0.05% of  $\alpha$ -tocopherol. Also, supercritical fluid extract at 200 bar contains 0.02% of carvacrol and 0.27% of  $\alpha$ -tocopherol. These antioxidant compounds might play vital role in contributing antioxidant properties of *Valeriana wallichii* supercritical fluid extracts. It has been reported that most natural antioxidative compounds often work synergistically with each other to produce a broad spectrum of antioxidative activities that creates an effective defence system against free radical attack [41].

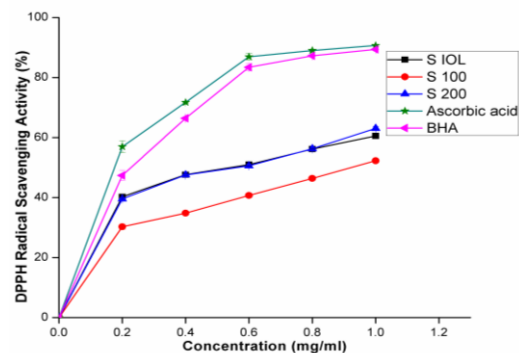


Fig. 1 Effect of *Valeriana wallichii* by DPPH radical scavenging activity in different concentration. Each value represents a mean  $\pm$  SD ( $p < 0.05$ )

### 3.3 Superoxide Radical Scavenging Activity

Percent superoxide radical scavenging capability of hydrodistilled *Valeriana wallichii* essential oil along with supercritical fluid CO<sub>2</sub> against the concentration was illustrated in Fig. 2. The superoxide scavenging effect of the essential oil (16.87% - 67.01%) supercritical fluid extract at 100 bar (23.60% - 70.64%) and supercritical fluid extract at 200 bar (18.17% - 64.05%) on superoxide radical linearly increased with increasing concentration from 0.05 to 1.0 mg. Supercritical fluid extracts and *V. wallichii* essential oil showed comparatively less percent scavenging activity in comparison with Ascorbic acid (46.53% - 80.51%) and BHA (42.37% - 80.33%). At the lower concentration (0.2 mg), all the extracts exhibited lesser radical scavenging effect than positive controls, however at 1 mg level, which exhibited more than 60% radical scavenging effect. Based on IC<sub>50</sub> value, the superoxide radical scavenging activity can be ranked in the following order, SFE of *V. wallichii* 100 bar (0.19 mg/mL) > SFE of *V. wallichii* 200 bar (0.30 mg/mL) > Hydrodistilled *V. wallichii* essential oil (0.33 mg/mL).

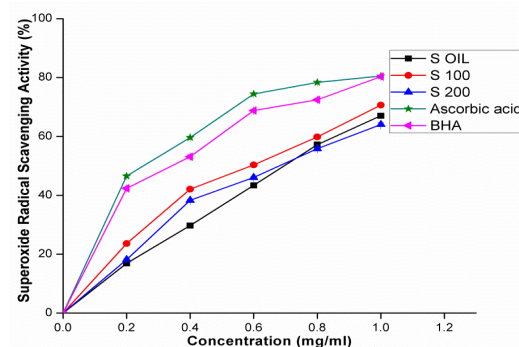
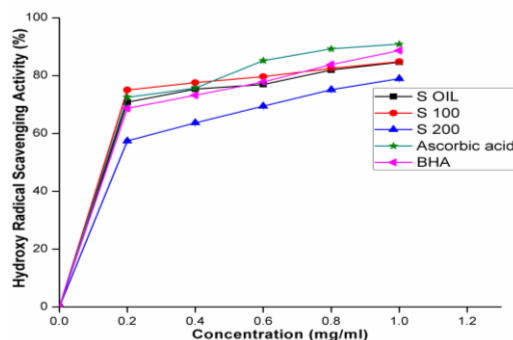


Fig. 2 Effect of *Valeriana wallichii* by superoxide radical scavenging activity in different concentration. Each value represents a mean  $\pm$  SD ( $p < 0.05$ )

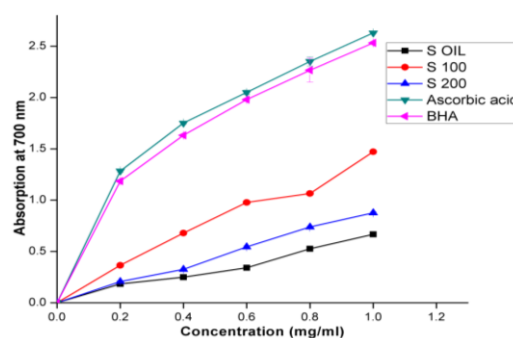
### 3.4 Hydroxyl Radical Scavenging Activity

Percent hydroxide radical scavenging capability of hydrodistilled *Valeriana wallichii* essential oil along with supercritical fluid CO<sub>2</sub> against the concentration was illustrated in Fig. 3. The scavenging effect of the essential oil (70.80% - 84.64%) supercritical fluid extract at 100 bar (75.03% - 84.83%) and supercritical fluid extract at 200 bar (57.40% - 78.95%) on hydroxide radical linearly increased with increasing concentration from 0.2 to 1.0 mg. All the essential oil and supercritical fluid extract showed good to excellent percent scavenging activity in comparison with BHA (68.65 - 88.69%) and ascorbic acid (72.52 - 90.89). At the lower concentration (0.2 mg) *V. wallichii* supercritical fluid extract

at 100 bar and essential oil are able to comparatively similar hydroxyl radical scavenging activity in comparison with ascorbic acid. All the tested extracts gave higher hydroxyl radical scavenging activity at 1.0 mg level which is relatively similar to positive controls. Based on IC<sub>50</sub> value, the percent hydroxyl radical scavenging activity can be ranked in the following order, SFE of *V. wallichii* 100 bar (0.13 mg/mL) > Hydrodistilled *V. wallichii* essential oil (0.14 mg/mL) > SFE of *V. wallichii* 200 bar (0.17 mg/mL). In the hydroxyl radical scavenging system, the concentration of hydrogen peroxide in water depends upon the phenolic compounds. Phenolic compounds present in the *V. wallichii* essential oil and supercritical fluid extract may be good electron donors and they may accelerate the conversion of H<sub>2</sub>O<sub>2</sub> to H<sub>2</sub>O [42].



**Fig. 3** Effect of *Valeriana wallichii* by hydroxy radical scavenging activity in different concentration. Each value represents a mean  $\pm$  SD ( $p < 0.05$ )



**Fig. 4** Effect of *Valeriana wallichii* by reducing power method in different concentration. Each value represents a mean  $\pm$  SD ( $p < 0.05$ )

### 3.5 Reducing Power

Reducing power of hydrodistilled *Valeriana wallichii* essential oil along with supercritical fluid CO<sub>2</sub> against the concentration was illustrated in Fig. 4. The increase of absorbance value at 700 nm denotes the increase of reducing power of particular sample. The reducing power of the essential oil (0.18 – 0.67) supercritical fluid extract at 100 bar (0.36 – 1.47) and supercritical fluid extract at 200 bar (0.21 – 0.88) linearly increased with increasing concentration from 0.2 to 1.0 mg. Supercritical fluid extracts comparatively showed good percent scavenging activity than essential oil. However, effects of extract as well as oil were lesser than ascorbic acid (1.2 – 2.6) and BHA (1.18–2.53). At the higher concentration (2 mg) *V. wallichii* supercritical fluid extract at 100 and 200 bar are able to similar reducing power as like lower concentration of positive controls. Based on IC<sub>50</sub> value, the percent hydroxyl radical scavenging activity can be ranked in the following order, SFE of *V. wallichii* 100 bar (0.28 mg/mL) > SFE of *V. wallichii* 200 bar (0.55 mg/mL) > Hydrodistilled *V. wallichii* essential oil (0.77 mg/mL). The reducing powers of essential oil might be due to their hydrogen-donating ability [43] and is generally associated with presence of reductones [44].

The components present in essential oil and supercritical fluid extracts of *V. wallichii* could act as good reductants, which could react with free radicals to stabilize and terminate radical chain reactions.

### 4. Conclusion

Summarizing these results, it can be concluded that the *Valeriana wallichii* DC essential oil and its supercritical fluid extracts exhibited broad spectrum of antioxidant activity against the tested various antioxidant assays. More studies are needed to clarify the antioxidant mechanisms of the antioxidant activity of essential oil and supercritical fluid extracts. On

the basis of above results, it was observed that *Valeriana wallichii* essential oil and supercritical fluid extracts provided comparatively equivalent antioxidative activity as compared to synthetic antioxidants, which provides a way to screen antioxidants for foods, cosmetics and medicine.

### Acknowledgement

We are thankful to Head, Chemistry Department, Gandhigram Rural University, Gandhigram for providing laboratory facilities and MR. P.C. Jain, Pragathi Aroma Oil Distillers Pvt. Ltd. for providing instrumentation facility for GC-MS analysis.

### References

- [1] African pharmacopoeia, Lagos, Organization of African Unity Technical and Research Commission, Africa, 1985.
- [2] J.S. Akshay, T. Shrikant, R.S. Mahesh, T.D. Sitaram, Pharmacognostic account of roots of *Valeriana wallichii* DC, Int. J. Pharm. Clinical Res. 4(4) (2012) 41-43.
- [3] R. Strachey, Catalogue of the plants of Kumaon and of the adjacent portions of Garhwal and Tibet, Periodical Experts, New Delhi, 1918, 84.
- [4] S. Fazal, K. Nasiara, M. Ibrar, Anti-inflammatory activity of methanolic and aqueous extracts of *Valeriana wallichii* DC rhizome, Pak. J. Plant Sci. 13(2) (2007) 103-108.
- [5] H.W. Youngken, Text book of pharmacognosy, Philadelphia, Blakiston, 1950.
- [6] R.D. Gaur, Flora of the District Garhwal North West Himalaya: With Ethnobotanical Notes, Trans Media, Srinagar, Uttarakhand, India, 1999.
- [7] R.N. Chopra, S.L. Nayar, I.C. Chopra, Glossary of Indian Medicinal Plants, Council of Scientific and Industrial Research, New Delhi, India, 1986.
- [8] N. Singh, A.P. Gupta, B. Singh, V.K. Kaul, Quantification of valeric acid in *Valeriana jatamansi* and *Valeriana officinalis* by HPTLC, Chromatographia 63(3-4) (2006) 209-213.
- [9] M. Howard, Traditional Folk Remedies, A comprehensive Herbal, Century paperbacks, London, UK, 1987.
- [10] A.H. Gilani, A. Khan, Q. Jabeen, F. Subhan, R. Ghafar, Antispasmodic and blood pressure lowering effects of *Valeriana wallichii* are mediated through K<sup>+</sup> channel activation, J. Ethnopharm. 100(3) (2005) 347-352.
- [11] J. Das, A.A. Mao, P.J. Handique, Terpenoid compositions and antioxidant activities of two Indian valerian oils from the Khasi Hills of north-east India, Nat. Prod. Comm. 6(1) (2011) 129-132.
- [12] V.K. Dua, M.F. Alam, A.C. Pandey, S. Rai, A.K. Chopra, V.K. Kaul, A.P. Dash, Insecticidal activity of *Valeriana jatamansi* (Valerianaceae) against mosquitoes, J. Am. Mosquito Control Assoc. 24(2) (2008) 315-318.
- [13] Z. Yan, T. Zhang, T. Xiao, L. Pan, J. Qin, Z. Zhang, C. Zuo, Antianxiety effect of *Valeriana jatamansi* Jones extract via regulation of the hypothalamus pituitaryadrenal axis, Neural Regen. Res. 5(14) (2010) 1071-1075.
- [14] N. Murakami, Y. Ying, M. Kawanishi, S. Aoki, N. Kudo, M. Yoshida, E.E. Nakayama, T. Shioda, M. Kobayashi, New rev-transport inhibitor with anti-HIV activity from *Valeriana Radix*, Bioorg. Med. Chem. Lett. 12(20) (2002) 2807-2810.
- [15] A. Khan, A.H. Gilani, Antidiarrhoeal and bronchodilatory potential of *Valeriana wallichii*, Nat. Prod. Res. 26(11) (2012) 1045-1049.
- [16] R. Bos, H. Hendricks, J.J.C. Scheffer, H.J. Woerdebad, Cytotoxic potential of valeriana constituents, and valerian tincture, Phytomed. 5 (1998) 219-225.
- [17] S. Suri, T.S. Thind, Antibacterial activity of some essential oils, Ind. Drugs Pharmaceut. Indus. 13 (1978) 25-28.
- [18] Y.P. Tang, X. Liu, B. Yu, Two new flavone glycosides from *Valeriana jatamansi*, J. Asian Nat. Prod. Res. 5(4) (2003) 257-261.
- [19] P.W. Thies, Linarinisovalerianate, a currently unknown flavonoid from *Valeriana wallichii*, Planta Medica. 16 (1968) 363-371.
- [20] Y.G. Chen, L.L. Yu, R. Huang, Y.P. Lv, S.H. Gui, 11-Methoxyviburtinal, a new iridoid from *Valeriana jatamansi*, Arch. Pharm. Res. 28(10) (2005) 1161-1163.
- [21] S. Lin, T. Chen, X.H. Liu, Y.H. Shen, H.L. Li, L. Shan, R.H. Liu, Iridoids and lignans from *Valeriana jatamansi*, J. Nat. Prod. 73(4) (2010) 632-638.
- [22] J. Xu, P. Zhao, Y. Guo, C. Xie, D.Q. Jin, Y. Ma, W. Hou, Iridoids from the roots of *Valeriana jatamansi* and their neuroprotective effects, Fitoterapia. 82(7) (2011) 1133-1136.
- [23] S. Palkhwal Sah, C.S. Mathela, K. Chopra, Elucidation of possible mechanism of analgesic action of *Valeriana wallichii* DC chemotype (patchouli alcohol) in experimental animal models, Ind. Jour. Exp. Biol. 48 (2010) 289-293.
- [24] S.P. Sah, C.S. Mathela, K. Chopra, *Valeriana wallichii* DC (Maaliol Chemotype): Antinociceptive Studies on, Experimental animal models and possible mechanism of action, Pharmacologia 3(9) (2012) 432-437.
- [25] S. Chemat, A. Lagha, H. Amar, P.V. Bartels, F. Chemat, Comparison of conventional and ultrasound - extraction of carvone and limonene from caraway seeds, Flav. Frag. Jour. 19 (2004) 188-195.
- [26] C.L. Wang, T. Weller, Recent advances in extraction of nutraceuticals from plants, Trend. Food Sci. Tech. 17 (2006) 300-312.
- [27] R.P. Adams, Identification of essential oil components by gas chromatography/mass spectrometry, Allured Publ. Corp, Carol Stream, IL. 4 (2007) Page number.
- [28] J.C.M. Barreira, I.C.F.R. Ferreira, M.B.P.P. Oliveira, J.A. Pereira, Antioxidant activities of the extracts from chestnut flower, leaf, skins and fruit, Food chem. 107 (2008) 1106-1113.
- [29] F. Liu, V. Ooi, S.T. Chang, Free radical scavenging activities of mushroom polysaccharide extracts, Life sci. 60 (1997) 763-771.
- [30] B. Halliwell, J.M. Gutteridge, C.E. Cross, Free radicals, antioxidants and human disease: where are now?, J. Labor. Clinic. Med. 119 (1992) 598-620.

- [31] M. Oyaizu, Studies on product of browning reaction: Antioxidative activities of browning products of browning reaction prepared from glucose amine, Jap. J. Nutri. 44 (1986) 307-315.
- [32] A. Kathirvel, V. Sujatha, In vitro assessment of antioxidant and antibacterial properties of Terminalia chebula Retz, Leaves, Asian Pacific J. Trop. Biomed. 2(2) (2012) S788-S795.
- [33] R.B. Arora, C. Arora, Hypotensive and tranquillizing activity of jatamansone (valeranone) a sesquiterpene from *Nardostachys jatamansi* DC, In; KK Chen, B Mukerji, Pharmacology of Oriental Plants, Pergamon Press, Oxford, Country, 1963, pp.51-60.
- [34] C.P. Khare, Indian Medicinal Plants: An illustrated Dictionary, Springer Publisher, US, 2007.
- [35] C.S. Mathela, M. Tewari, S.S. Sammal, C.S. Chanotiya, *Valeriana wallichii* DC, a new chemotype from Northwestern Himalaya, J. Essential Oil Res. 17 (2005) 672-675.
- [36] C.S. Mathela, R.C. Padalia, C.S. Chanotiya, Kanokonylacetate-rich Indian valerian from Northwestern Himalaya, Nat. Prod. Comm. 4 (2009) 1253-1256.
- [37] Y.P. Tang, X. Liu, Y. Biao, Two new flavone glycosides from *Valeriana jatamansi*, J. Asian Nat. Prod. Res. 5 (2003) 257-261.
- [38] A. Pandey, Y.N. Shukla, Naphthoic acid derivative from *Valeriana wallichii*, Phytochem. 32 (1993) 1350-1359.
- [39] A. Panday, G.C. Uniyal, Y.N. Shukla, Determination of chemical constituent of *Valeriana wallichii* by reverse phase HPLC plus, Ind. Jour. Pharmaceut. Sci. 56 (1994) 56-58.
- [40] M.F. Alam, A.C. Pandey, S. Rai, A.K. Chopra, V.K. Kaul, A.P. Dash, Insecticidal activity of *Valeriana wallichii* against mosquitoes, J. Am. Mosquito Control Assoc. 24 (2008) 315-318.
- [41] F. Lu, L.Y. Foo, Antioxidant activities of polyphenols from sage (*Salvia officinalis*), Food Chem. 75 (2001) 197-202.
- [42] R.T. Ruch, S.U. Chung, J.E. Kalaunig, Spin trapping of superoxide and hydroxyl radicals, Method. Enzymol. 105 (1984) 198-209.
- [43] P.D. Duh, Antioxidant activity of Budrock (*Arctium laooa* Linn): Its scavenging effect on free radical and active oxygen, Jour. Am. Oil Chem. Soc. 75 (1998) 455-461.
- [44] C. Hyang-Sook, H.S. Song, H. Ukeda, M. Sawamura, Radical-scavenging activities of citrus essential oils and their components: detection using 1,1-diphenyl-2-picrylhydrazyl, J. Agri. Food Chem. 48 (2000) 4156-4161.